First Report of *Phytophthora insolita* and *P. inflata* on Rhododendron in Ohio. Antonino Testa, Institute of Biochemical Biotechnology—Programma Miur "Rientro dei Cervelli", Università Politecnica delle Marche, Ancona, Italy; Mikael Schilb, Department of Life Science, Otterbein College, Westerville, Ohio and Department of Plant Pathology, The Ohio State University, Columbus; Jeffrey S. Lehman, Department of Life Science, Otterbein College, Westerville, Ohio; Gennaro Cristinzio, Department of Arboricoltura, Botany, and Plant Pathology, Università di Napoli "Federico II", Portici (NA), Italy; and Pierluigi Bonello, Department of Plant Pathology, The Ohio State University, Columbus. Plant Dis. 89:1128, 2005; published on-line as DOI: 10.1094/PD-89-1128B. Accepted for publication 11 July 2005.

During August 2003, we conducted a statewide survey of rhododendrons to determine if Phytophthora ramorum was present in Ohio ornamental nurseries. In total, 240 samples were randomly collected in 12 nurseries throughout Ohio from rhododendrons showing foliar necrotic lesions and twig dieback symptoms. The samples yielded 51 Phytophthora spp. isolates on PARP-V8 agar. The internal transcribed spacer (ITS) region of all isolates was amplified using the universal primers ITS1 and ITS4 and was sequenced. Consensus sequences from sense and antisense were then blasted against the GenBank database, allowing for the identification to species of ~80% of all isolates. These identifications, and the ~20% unknowns, were confirmed using blind morphological tests on the basis of the following parameters: colony morphology; shape and dimensions of sporangia and type of papillae; dimensions of oogonia and oospores; type and position of antheridia; presence or absence of chlamydospores; presence or absence and morphology of hyphal swellings; and growth rate at 35°C according to the Revisited Tabular Key of the species of *Phytophthora* (1). No *P. ramorum* was detected among the isolates; however, *P.* cactorum, P. citricola, P. citrophthora, and P. nicotianae were detected. We also found two occurrences of *P. inflata* Caros & Tucker and one of *P. insolita* Ann & Ko. (*P. inflata: e-value <* $e(^{-179})$, identities >95%; P. insolita: e-value = 0.0; identities = 95%.) P. inflata was isolated from two tissue types, a dead twig and a necrotic leaf tip. P. insolita was isolated from a necrotic leaf tip. Identity of the two species was confirmed morphologically using the parameters listed above as well as the following measurements (N = 40; all in μ m) (1): P. inflata – sporangia: 40×24 ([24 to $(68] \times [18 \text{ to } 34]);$ oogonia: 34.6 (28 to 40); oospores: 30.8 (25 to 38); *P. insolita* – sporangia: 42×10^{-10} 28 ([34 to 56] × [22 to 38]); oogonia: 32 (26 to 36); oospores: 26 (22 to 30). Koch's postulates were satisfied by inoculating two rhododendron plants (cvs. PJM and Nova Zembla) with the putative pathogens. On each plant, each of three leaves was pierced with a dissecting needle and was inoculated by placing a 0.5-cm-diameter plug of mycelium that was taken from the margin of a colony actively growing on PARP-V8 agar on the wound. The inoculum was retained using clear adhesive tape. A similar procedure was used for twigs. Controls consisted of inoculations with sterile PARP-V8 agar medium. Both cultures of P. inflata and P. insolita produced necrotic lesions in all inoculations on both tissue types within 1 week, and they were reisolated from the margins of lesions on PARP-V8. The lesion margin was at least 2 cm away from the inoculum plug in leaf inoculations and several centimeters in twig inoculations. To our knowledge, this is the first report of P. inflata and P. insolita occurring on rhododendron and the first time P. insolita has been reported outside of Southeast Asia where it has been recovered only from soil.

Reference: (1) D. J. Stamps et al. Mycol. Pap. No. 162. CAB Int. Mycol. Inst. Wallingford, UK, 1990.